

CALLUS INDUCTION, SOMATIC EMBRYOID FORMATION AND PLANT REGENERATION IN COTTON (*GOSSYPIUM HIRSUTUM* L.)

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High frequency of callus induction and somatic embryogenesis was observed on MS medium containing various concentrations and combinations of different growth regulators. Among the various explants, young leaf was found to be best for maximum frequency of callus induction on MS medium fortified with NAA (2.0 mg/l) in combination with 2 iP (3.0 mg/l). These calli developed embryoids on MS medium containing 2 iP and ABA (2.0 mg/l each) with 10 mM glutamine. Embryoids formed here developed into plantlets and plant regeneration frequency was low.

Keywords: Callus culture; Cotton; Explants; Hormones; Plant regeneration; Somatic embryogenesis.

Introduction

Tissue culture of cotton has been notoriously difficult¹. With the first report of regeneration of cotton plants², progress in plant regeneration has not been achieved rapidly, because of the lengthy culture period³. The plant regeneration was achieved from hypocotyl explants of American Coker 201 and 315 cultivars. Recently, various types of embryoids were obtained and developed into fertile plants⁴. The objective of this investigation was to devise a tissue culture method for plant regeneration via somatic embryogenesis to our Indian cultivar MCU-7.

Materials and methods

Delinted seeds were surface sterilized with 0.1% (W/V) mercuric chloride for 5 min and germinated on MS⁵ basal medium under dark. Different explants viz., hypocotyl, cotyledon young leaf and immature embryo were used for callus initiation. The MS medium supplemented with two auxins (2, 4-D and NAA) along with three different cytokinins (Kin, 2 iP and BAP) were used.

Cut pieces of explants were placed in

25 x 150 mm culture tubes with 20 ml of the medium containing respective growth regulator combinations and incubated at 25±2 C with 16 hrs photoperiod in cool-white fluorescent light (1500 lux). Callusing percentage was calculated after 20 days. MS medium supplemented with different levels of auxins and cytokinins and other organic supplements like ABA and glutamine were used for regeneration study.

Results and Discussion

Callus induction and proliferation: In the present study, different explants were used for callus initiation. NAA (2.0 mg/l) along with 2 iP (3.0 mg/l) was found to be the best combination for callus induction (Fig 1). However, the combination and dosage varied with explants.

The leaves had maximum callus induction in NAA (2.0 mg/l) along with 2 iP (3.0 mg/l). In hypocotyl, NAA (2.0 mg/l) in combination with Kin (0.5 mg/l) induced the maximum frequency of callus. The highest frequency of callus induction was observed on MS medium containing 2, 4-D and Kin

Table 1. Optimum dosage of auxin and cytokinin for maximum frequency of callus induction from different explants of cotton.

Growth Hormones	EXPLANTS							
	Hypocotyl		Cotyledon		Young leaf		Immature embryo	
2,4-D : Kin	0.1	0.5 (75.3)	0.1	0.1 (83.3)	0.5	0.5 (80.3)	0.1	1.0 (60.0)
2,4-D : 2 iP	0.1	2.0 (69.7)	0.1	3.0 (75.3)	0.5	2.0 (71.7)	0.1	1.0 (62.0)
2,4-D : BAP	0.1	1.5 (65.3)	0.1	1.5 (54.3)	0.3	2.0 (66.0)	0.1	2.0 (46.7)
NAA : Kin	2.0	0.5 (82.7)	1.0	0.5 (72.7)	2.0	0.5 (78.0)	2.0	0.5 (63.0)
NAA : 2 iP	1.0	2.0 (71.7)	1.5	2.0 (75.3)	2.0	3.0 (85.7)	0.5	2.0 (55.3)
NAA : BAP	1.0	1.5 (52.7)	1.5	1.0 (54.7)	2.0	2.0 (68.7)	1.0	1.0 (38.7)

Figures in parenthesis are indicated percent of callus induction

(0.1 mg/l each) for cotyledon explant whereas 2,4-D (0.1 mg/l) and Kin (1.0 mg/l) combination was found to be best for maximum frequency of callus induction for immature embryos.

The differential response of the explants towards callus induction reveals the influence of explants, growth regulators and their concentrations. In the present study, both auxins had a similar effect on callus induction. However, the two auxins had individual effect when used along with different cytokinins. This apparent difference in callus induction with different explants might be due to the presence of endogenous auxin.

When 2, 4-D was used as auxin along with 3 different cytokinins for callus induction, maximum frequency of callus induction was in cotyledon (83.3%) with 2, 4-D and BAP (0.3 and 2.0 mg/l, respectively). A dose of 2, 4-D and 2 iP (0.1 and 3.0 mg/l, respectively)

was also the best combination for cotyledon (Table 1).

In the differential performance of 2, 4-D no callus induction was observed using cotyledon explant¹. This contradicts to the present results where maximum callus induction (83.3%) with cotyledon was recorded with 2, 4-D as auxin (Table 1).

When NAA was used as auxin along with 3 different cytokinins, NAA with 2 iP combination induced highest frequency of callus induction (85.7%) at (2.0 and 3.0 mg/l respectively) for leaf tissue, whereas in hypocotyl, NAA (2.0 mg/l) and Kin (0.5 mg/l) combination found to be best for maximum frequency of callus induction. NAA and BAP (2.0 mg/l each) combination was also found to be better for maximum frequency of callusing in young leaves. Trolinder and Xhixian⁶ also reported that the NAA and 2 iP combination found to be the best combination for callusing in cotton.

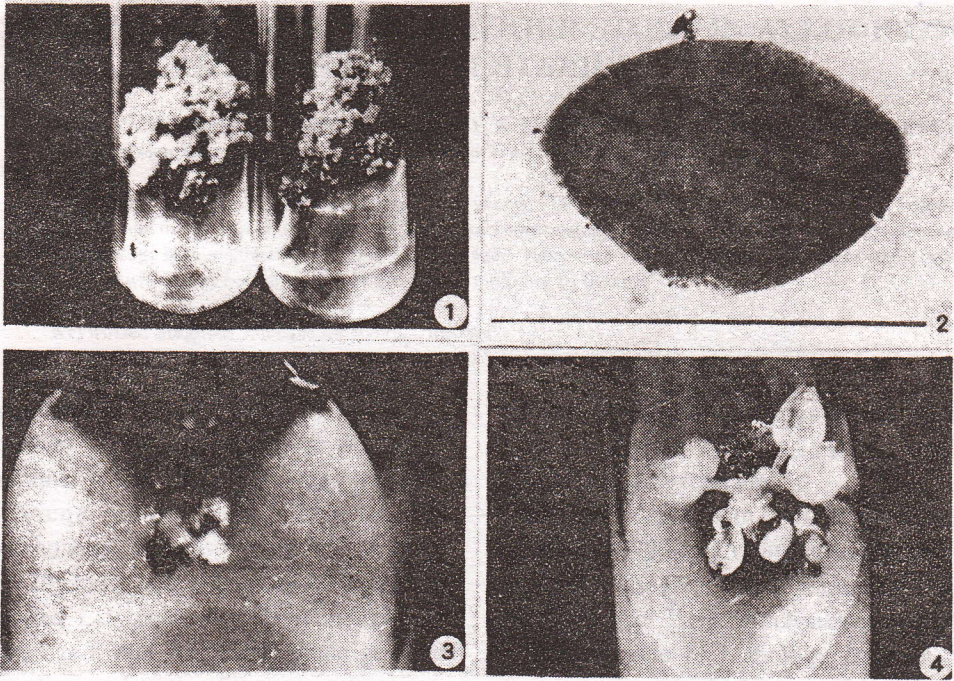


Fig. 1-4 : Efficient callus induction; (2) Heart shaped embryoid; (3) Plantlet development; (4) Well developed plantlet.

Regeneration: In the present study, MS medium supplemented with 2 iP and, ABA (1, 2 and 3 mg/1 each), glutamine (5 to 20 mM) along with organic additives were tried (Data not shown). Addition of ABA and 2 iP (2.0 mg/1 each) with 10 mM glutamine resulted in the embryoid formation (Fig 2). MS medium supplemented with ABA (5 μ m) produced six types of somatic embryos in cotton⁴.

Increased concentration of KNO_3 and addition of $MgCl_2$ was not found to be significant for embryoid formation. High frequency of somatic embryos was obtained with the addition of 2x. KNO_3 and $MgCl_2$ ^{1,3,7,8}.

Plant recovery: In the present study, embryos were transferred to MS medium with the elimination NH_4NO_3 but with 2x KNO_3 and $MgCl_2$ (750mg/1), solidified in 2% (W/V) gelrite. Embryoids developed into leaf

structures and developed into small plantlets (Fig. 3 &4). Cousin *et al.*⁹ used the same medium for germinating Australian cultivar siokera 1-3 with less frequency.

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