

SALT STRESS INDUCED CHANGES IN GROWTH, PIGMENT, PROLINE AND $\text{Na}^+ - \text{K}^+$ CONTENT IN *ORYZA SATIVA* L.

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Salt stress reduced the germination rate of rice (*Oryza sativa* L.) seeds. An uniform decrease in root length was marked in case of both NaCl and CaCl_2 treatments compared to shoot growth except for a minor changes in NaCl treatment. An uniform decrease in fresh and dry matter content was seen for both the salts. An increase in Na^+ uptake was recorded with a decrease in K^+ content in the rice shoot. Salt stressed seedlings showed a decrease in chlorophyll and carotenoid content from control seedlings. For both the salts tested an uniform accumulation of proline was visible.

Keywords : Growth; K^+ ; Na^+ ; *Oryza sativa*; Pigment; Proline.

Introduction

Salt stress is an abiotic environmental stress that limits the growth and development of plants which can be seen through changes in plant's morphology, physiology and metabolism. Rice is a sub-tropical cereal crop which is generally considered as salt sensitive than later stage^{1,2}. Salt stress is known to affect germination, growth, pigment content, mineral composition, ionic balance, proline accumulation and enzyme activities in various plants³⁻⁹. The major objective of the present study was to investigate the salt stress induced changes in germination, growth, pigment, proline content and $\text{Na}^+ - \text{K}^+$ accumulation in a salt sensitive traditional rice variety.

Materials and Methods

Dry graded uniform rice (*Oryza sativa* L. var. Beeroin) seeds were procured from Assam Agricultural University's Regional Rice Research Station, Karimganj, Assam and were surface sterilised with 0.1% mercuric chloride (w/v) for 10 min followed by three rinses in sterile distilled water. The seeds were germinated in petriplates containing Whatman No. 1 filter paper moistened with distilled water and under different (0, 0.5, 1, 2 and 3%) salt solutions and keep in BOD incubator at $25 \pm 2^\circ\text{C}$.

On the 3rd day germination rate was calculated for both the salts (treated and untreated). The germinated seeds (untreated) were transferred to plastic

glasses containing Yoshida solution and kept in a growth chamber under continuous white light. White light was provided by cool fluorescent white tubes (36 W Philips TLD) with a photon flux density of $52 \mu\text{Em}^{-2}\text{s}^{-1}$ (PAR) under a 16-h photoperiod. On the 12th day, rice seedlings growing in Yoshida solution were given a treatment of NaCl and CaCl_2 (0, 0.5, 1, 2 and 3%) solution for 4h. After 4h, seedlings were rehydrated with fresh Yoshida solution and after 24 h i.e., on 13th day plants were taken. For different growth parameters, 13 days old seedlings growing in Yoshida solution alone and with various concentrations of NaCl and CaCl_2 were studied for root and shoot growth in terms of length. Triplet samples of uniform seedlings washed with the help of electronic balance and measurements were noted. Seedlings were kept in an oven at 80°C for 72h and dry matter was determined. Both fresh and dry masses were expressed as biomass produced in mg/seedlings. Primary leaves and roots were sampled, oven dried and acid digested as per the methods of Humpries¹⁰. Na^+ and K^+ were estimated from the samples using a Flame Photometer (Systronics - India).

Analysis of pigments and metabolites was done on 13 days old seedlings growing in Yoshida solution and under different salt treatments. For the analysis of chlorophyll, carotenoid and proline, primary leaves of the seedlings were taken. The extraction of chlorophyll and carotenoid was done using

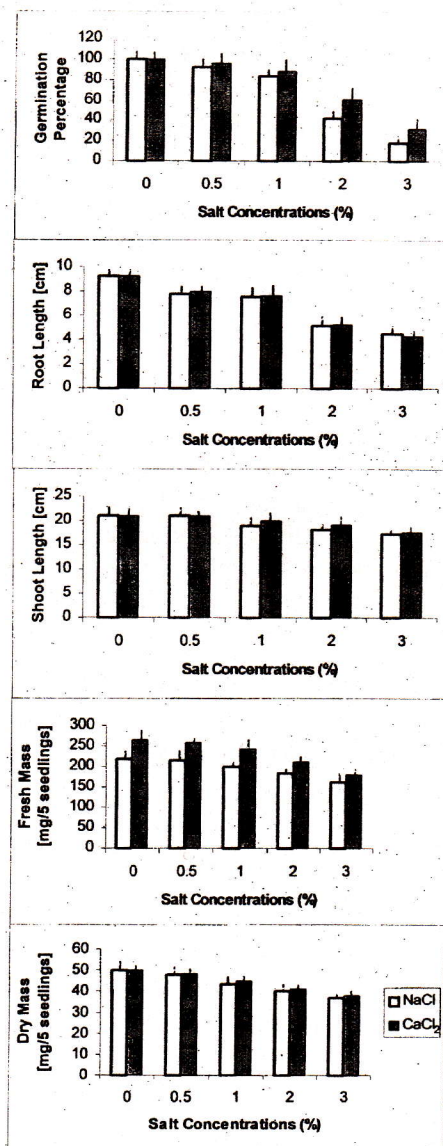


Fig. 1. Changes in germination percentage, root length, shoot length, fresh mass and dry mass of 13 days old rice seedlings subjected to salt treatments. Data presented are mean \pm SE.

80% cold alkaline acetone¹¹. The chlorophyll and carotenoid content was expressed as $\mu\text{g/g}$ fresh weight of sample. Primary leaves (both treated and untreated) were homogenised with 3% aqueous sulfosalicylic acid and centrifuged at 3,000 g for 10 min. Proline was estimated as per method of Bates *et al*¹². Each experiment

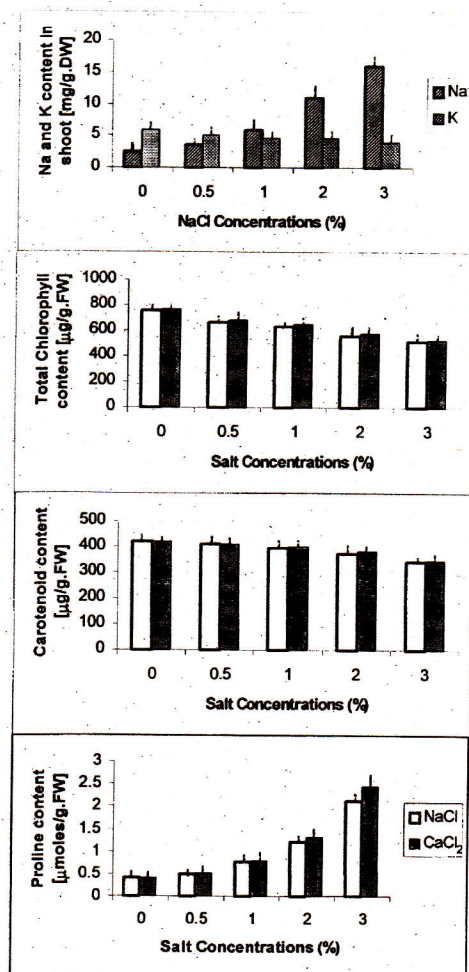


Fig. 2. Changes in Na^+ - K^+ content, total chlorophyll, carotenoid and proline content of 13 days old rice seedlings subjected to salt treatment. Others same as Fig. 1.

was done in triplicates and data represent mean \pm SE.

Results and Discussion

There is a gradual decrease in the germination rate with the increase in the salt concentration upto 1.0% compared to control and less than 50% has been observed

in 2.0% NaCl. 3.0% showed a maximum toxicity where almost 80% seeds could not germinate. In case of CaCl_2 treated samples, similar results were observed with less toxicity as compared to NaCl treated rice seeds. LC_{50} in NaCl is 2.0% and LC_{50} in CaCl_2 is 3.0% treated rice seeds was observed. Results showed a uniform decrease in seed germination rate with a maximum in NaCl as compared to CaCl_2 . Similar results for various other plants have been reported by several other workers^{9,13-15}. There is a gradual decrease in the root length of the rice plants subjected treated with NaCl and CaCl_2 . The effect of the salts on the roots is toxic where the length of the root has been reduced to almost half compared to control. However, the CaCl_2 treated samples were affected more than that of NaCl treated rice plants. Root length decreased in both the salts whereas a slight increase in shoot growth was recorded. There is a decrease in 0.5% in the shoot length of NaCl treated plants compared to control followed by a gradual increase with the increase in the salt concentration. Similar result was observed in CaCl_2 treated rice plants but more severely in 3.0% concentration (Fig. 1). A gradual increase in the dry weight with the increase in the salt concentration was observed in 13 days old NaCl treated plants. A similar result was observed in CaCl_2 treated plants. The dry weight was reduced to almost one fourth of the fresh weight. The effect of salt stress on the performance and growth are mediated through decrease in stimulating conduction and photosynthesis. It is known that the degree to which the growth is reduced by salinity differs greatly with species and then cultivar within a species¹⁶. Fresh and dry masses though decreased with a slight increase in some of the concentrations showing a modulatory role as reported elsewhere¹⁷.

Figure 2 illustrates the changes in the Na^+ and K^+ content in the shoot of 13 d old rice seedling under different concentration of NaCl. There is an increase in the Na^+ ion content with the increase in the salt concentration with the maximum of at 3.0%.

The K^+ content decreased with the increase in the NaCl concentration compared to control. The increase in Na^+ content with a decrease in K^+ content in shoot tissues suggested a better Na^+ uptake which disturbed ionic balance as seen for other salt sensitive rice varieties¹⁸⁻²⁰. Higher salinity disturbs $\text{Na}^+ : \text{K}^+$ ratio in the plant, which impairs the proteins metabolism of the plant. It has been reported that varieties of rice avoid the sodium toxicity by better potassium levels in the tissue but the results showed a relative decrease along the salinity gradient^{21,22} proving the cultivar to be a salt sensitive one. The NaCl treated leaf tissues showed a decrease in the chlorophyll and carotenoid content with the increase in the salt concentration. 1.0% NaCl treated plants showed a sharp increase followed by a decrease in 2.0% and 3.0%. Similar results were seen for CaCl_2 treated seedling. Total chlorophyll showed a uniform decrease in NaCl and CaCl_2 treatments with the increasing concentrations for seedling of both the ages. Similar reports are there for various other plants^{6,23-24}. Carotenoid content decreased uniformly in salt stressed plants, suggesting an inhibition of photosynthetic efficiency in salt sensitive rice plant²⁴. Decreased chlorophyll synthesis or chlorophyll molecules breakdown, rate of light reaction in photosynthesis, decreased rate of ribulose biphosphate carboxylase - oxygenase (RUBISCO) enzyme activity has been reported under salt stress²⁵. There is an uniform increase in the proline content compared to control in both the 13 and 14 days NaCl treated leaf tissues with a maximum in 3.0% NaCl. 14 days old plants contain higher proline than the 13 days old seedlings. Similar result was noticed in CaCl_2 treated rice leaves. An increase in proline content under both NaCl and CaCl_2 salinity suggested an osmoprotection to rice seedling under salt stress as reported for other plants as proline act as an osmoprotectant by providing non-toxic sinks for carbon and nitrogen preservation^{18,26,27}. Increasing proline level is considered to help the cells in

osmoprotection as well as in regulating the redox potential, scavenging hydroxy radicals in the protection against denaturation of various macromolecules²⁸.

References

1. Pearson G A 1959, *Soil Sci.* **87** 198
2. Yeo A R and Flowers T J 1982, *Physiol. Plant.* **56** 343
3. Greenway H and Munns R 1980, *Annu. Rev. Plant Physiol.* **31** 149
4. Hampson C R and Simpson G M 1990, *Can. J. Bot.* **68** 524
5. Yeo A R, Lee K S, Izard P, Boursier P J and Flowers T J 1991, *J. Expt. Bot.* **42** 881
6. Misra A N, Sahu S M, Mishra M, Singh P, Merra I, Das N, Kar M and Sahu P 1997, *Biol. Plant:* **39** (2) 257
7. Mathew R and Chandrashekar K R 1998, *J. Phytol. Res.* **11**(1) 23
8. Promila K and Kumar S 2000, *Biol. Plant.* **43**(3) 423
9. Dash M and Panda S K 2001, *Biol. Plant.* **44**(4) 587
10. Humphries E C 1956, *Modern Methods of Plant Analysis*. Eds. Peach K and Tracey N V Vol. 1. pp. 468 Springer - verlag, Berlin.
11. Arnon D I 1949, *Plant Physiol.* **24** 1
12. Bates L S, Waldren R P and Teare L D 1973, *Plant and Soil.* **39** 205
13. Zidan M A and Elewa M A 1995, *Ind. J. Plant Physiol.* **38**(1) 57
14. Hagar H, Ueda N and Shah S V 1996, *Am. J. Physiol.* **271** F209
15. Ozturk M, Baslar S, Dogan Y and Mert H H 1997, *Cruciferae Newsletter.* **19** 69
16. Shannon M C and Grieve C M 1999, *Scientia Hort.* **78** 5
17. Rascio A, Cedola M C, Sorrentino G, Pastore D and Wittoner G 1988, *Physiol. Plant.* **73** 122
18. Singh A K and Singh R A 1999, *Ind. J. Plant Physiol.* **4**(2) 111
19. Krishnamurthy R, Anabazhagan M and Bhagwat K A 1987, *Indian J. Plant Physiol.* **30** 183
20. Datta K S, Kumar A, Verma S K and Angrish R 1996, *Indian J. Plant Physiol.* **1** (NS) 102
21. Mass E V and Poss J A 1989, *Irrig. Sci.* **10** 29
22. Qadar A 1991, *Ind. J. Plant Physiol.* **34** 319
23. Khaveri-Nejad R A and Chaparzadeh N 1998, *Photosynthetica.* **35**(3) 461
24. Singh A K and Singh R A 1999, *Ind. J. Plant Physiol.* **4**(1) 49
25. Ingram J and Bertels D 1996, *Annu. Rev. Plant Physiol. Plant Mol. Biol.* **47** 377
26. Joyee P A, Aspinall D and Paleg L G 1992, *Aust. J. Plant Physiol.* **19** 249
27. Delauney A J and Verma D P S 1993, *Plant J.* **4** 215
28. Matysik J, Alia, Bhalu B and Mohanty P 2002, *Curr. Sci.* **82** 525