

VASE LIFE STUDIES ON VANDA 'JOHN CLUBB' INFLORESCENCES

P.G. LATHA and M. JAYASREE*

Tropical Botanic Garden and Research Institute, Palode, Thiruvananthapuram 695562, India.

*Mahatma Gandhi College, Thiruvananthapuram 695004, India.

Freshly cut inflorescences of the orchid hybrid, Vanda 'John Clubb', after a conditioning treatment were used to study their post-harvest vase life. Solutions containing silver nitrate significantly increased vase life. The hypothesis that ethylene production is the cause of the reduced vase life of Vanda 'John Clubb' is discussed.

Keyword : Ethylene inhibitor; silver nitrate; Vanda 'John Clubb'.

One of the main limitations of the floriculture industry is the post harvest loss during the transit from the field to the market. The use of floral preservatives is recommended during the entire marketing chain to preserve the quality of cut flowers after harvest¹. Cut flowers are often held in holding/vase solutions containing carbohydrate as a source of energy and a biocide to inhibit the growth of *micro-organisms*². Sucrose is the commonly used carbohydrate, being an excellent osmoticum and respiratory substrate³.

The major chemical pollutant affecting the senescence of cut flowers is ethylene⁴. Orchid blossoms including those of *Vanda* hybrids are extremely sensitive to ethylene⁵. A perusal of literature revealed that very few reports are available on the use of holding solutions to increase the vase life of hybrid Vandas. Hence the present study was undertaken.

Vanda 'John Clubb' (*V. Cooperi* x *tessellata* var. *rufescens*) is an attractive free flowering Sri Lankan orchid hybrid with mauve - pink flowers, having bright crimson spotted lip⁶.

Inflorescences of *Vanda* 'John Clubb' having all opened flowers except the terminal bud were randomly harvested in the morning (8 am), from the Orchidarium of the Institute. They were subjected to a conditioning treatment immediately after harvest by placing the lower end of the inflorescence stalk in tap water (pH 3.0) for 2 h as reported elsewhere⁷.

The conditioned inflorescences were trimmed (0.5 cm from the base) and subjected to the vase solution treatments. The vase solution (50 ml) was held in 250

ml boiling tubes. It contained 0.05% of the commercial fungicide, Dithane M-45 (Indofil Chemical Company, Mumbai) and 2% sucrose in distilled water, in addition to varying concentrations of silver nitrate (AgNO_3 - 10, 25 and 40 mg/l). Inflorescences, evaluated in a solution of 2% sucrose in distilled water served as control. The holding solution was replaced on every other day after trimming off 0.5 cm of the *inflorescence* stalk base, till the end of the experiment. All vase life evaluations were conducted at $28 - 30 \pm 2^\circ$ C and a relative humidity of $70 \pm 10\%$ in the laboratory. There were 5 replicates to each treatment. The average number of days taken for the appearance of wilting of the first flower (from below) was defined as the vase life of the *inflorescence*.

The longevity of the inflorescence was increased almost 3 - fold over the control by holding in vase solution containing 25 mg/l AgNO_3 . At the lower concentration used (10 mg/l), the vase life was only twice that observed in control. With higher concentration of AgNO_3 (40 mg/l), the vase life did not show further increase (Table 1).

The aims of post - harvest handling methods should be to reduce physico-chemical changes responsible for deterioration of the aesthetic value of the inflorescences. The visual changes during the post-harvest period are related to the age of the flowers, upsurge in ethylene (C_2H_4) evolution and accumulation of the ethylene precursor, 1-amino-cyclo-propane carboxylic acid (ACC)⁸. The major chemical pollutant affecting senescence is ethylene. Orchid flowers⁵ are particularly sensitive to ethylene which regulates the ageing^{5,9,10} and

Table 1. Effect of silver nitrate (AgNO₃) on the vase life of *Vanda* 'John Clubb' inflorescences.

Treatment	Mean Vase Life (Days)
DW + sucrose (2%) + Dithane M-45 (0.05%)	5.2 ± 0.1
DW + sucrose (2%) + Dithane M-45 (0.05%) + AgNO ₃ (10mg/l)	10.1 ± 0.2**
DW + sucrose (2%) + Dithane M-45 (0.05%) + AgNO ₃ (10mg/l)	15.0 ± 2.0**
DW + sucrose (2%) + Dithane M-45 (0.05%) + AgNO ₃ (10mg/l)	15.1 ± 2.2**

Values are the mean ± SD, n = 5, **P ≤ 0.001

senescing flowers produce their own ethylene¹¹. In the orchid inflorescence, the peak in ethylene evolution coincides with the end of its decorative role. For success in the orchid cut flower industry therefore, proper management of ethylene - like pollutants in the environment of flowers and regulation of C₂H₄ biosynthesis within the flowers is cogent to gear up their keeping quality.

Significant C₂H₄ biosynthesis has been reported from *Vanda* blossoms¹¹. In the present study, the significant increase in vase life of the *Vanda* inflorescences, held in vase solution containing AgNO₃ (25 mg/l) is perhaps due to the capacity of AgNO₃ to inhibit ethylene biosynthesis in flower tissue as has been reported elsewhere¹² and also because this concentration of AgNO₃ was optimal for C₂H₄ inhibition. Silver is known to delay the senescence of flowers including orchids⁸. Silver inhibits ethylene biosynthesis, possibly by binding to sulphhydryl groups or by acting competitively⁸. Silver thiosulphate has been reported to increase the vase life of *Dendrobium* 'Jacquelin Thomas' and *D.* 'Pompadour'. Perhaps, the carbohydrate used, sucrose was at the optimal concentration (2%) and helped to prolong the flower quality as reported elsewhere¹³. However very high concentrations of sugars in flowers do not necessarily prevent senescence. Similar studies with other ethylene inhibitors like aminoethoxyacetic acid (AOA) also gave concomitant results with other orchid blossoms¹⁴. The mild concentration of the fungicide, Dithane M-45 used in the study (0.05%) did not hinder the keeping quality of the flowers. At the same time, it helped to check growth of micro-organisms in the holding solution, Silver nitrate is also reported to have *microbicidal* properties in holding solutions¹⁵ and may also have contributed

to check growth of micro-organisms in the present study.

The present study thus revealed the importance of using ethylene inhibiting chemicals in the holding solution to increase the vase life of the orchid cut flower, *Vanda* 'John Clubb'

Acknowledgements

The authors are indebted to the Director, Tropical Botanic Garden and Research institute, Thiruvananthapuram and the Head, Department of Botany, Mahatma Gandhi College, Thiruvananthapuram, for laboratory facilities.

Reference

1. Kofranek AM and Halevy AH 1976, *Hort Science* 11 572
2. Reid MS and Kofranek AM 1980, *Chronicles of Horticulture* 20 25
3. Jomy TG, Sabina GT and Sreekandan Nair G 1999, *Journal of the Orchid Society of India* 131 41
4. Halevy AH and Mayak S 1979, In: *Horticultural Review*, J Janick (ed.) AVI Publishing, Connecticut, USA pp 59
5. Arditti J 1979, *Advances in Botanical Research* 7 621
6. Henderson MR and Addison GH 1969, In: *Malayan Orchid Hybrids*. Government Printing office, Singapore, 95
7. Jomy TG 1998, *Enhancement of post - harvest life of Dendrobium flower*. M.Sc. Hort. Thesis, Kerala Agricultural University, Thrissur, Kerala.
8. Nair H 1984, *Malayan Orchid Review* 18 62
9. Chadwick AV, Nyman CP, Arditti J 1986, *Lindleyana* 1 164
10. Hew CS, Gonk SS, Lin WS, Yong JWH 1995, *Lindleyana* 10 43
11. Burg SP and Dijkman MJ 1967, *Plant Physiology* 42 1648
12. Halevy AH and Kofranek AM 1977, *Journal of the American Society of Horticultural Science* 102 76
13. Pritchard MK and Hew CS 1998, *Lindleyana* 13 16
14. Porat R 1994, *Lindleyana* 9 85
15. Larsen FE and Cromarty RW 1967, *HortScience* 11 572